

PII: S0960-894X(96)00343-5

SYNTHESIS AND ACTIVITY OF STAUROSPORINE ANALOGS WITH A LACTONE FUNCTIONALITY

Rintaro Yamada, Koichiro Fukuda, Masashi Kawanishi, Yutaka Ohmori, Makoto Nasu, Minoru Seto, Yasuharu Sasaki

Institute of Life Science, Asahi Chemical Industry Co.,Ltd.
632-1 Mifuku Ohito-chou Tagata-gun Shizuoka 410-23, Japan

Toshiaki Sunazuka, Li Zhuorong, Nobuyuki Funato, Mieko Iguchi Yoshihiro Harigaya, Yuzuru Iwai, Satoshi Ōmura*

Research Center for Biological Function, The Kitasato Institute and School of Pharmaceutical Sciences, Kitasato University

5-9-1 Shirokane, Minato-ku, Tokyo 108, Japan

Abstract. Staurosporine analogs, whose γ -lactam functionality was converted into γ -lactone, were synthesized. The key step involves remarkably efficient ring-opening and then intramolecular cyclization reactions. In general, those compounds display potent anti-platelet aggregation activity and show higher selectivity against platelet as compared with smooth muscle contraction. Copyright © 1996 Elsevier Science Ltd

Staurosporine (1), a natural product was isolated by \overline{O} mura et al. 1 first from Saccharothrix staurosporeus and found almost a decade later to be a very potent inhibitor of protein kinase C (PKC) by Tamaoki et al. 2. However 1 has various pharmacological effects such as inhibition of platelet aggregation³, inhibition of smooth muscle contraction⁴, promotion of cell differentiation⁵ and growth inhibition of smooth muscle cells⁶, since it is completely unselective with respect to inhibition of other protein kinases (e.g.myosin light chain kinase, cyclic AMP dependent protein kinase, cdc2 kinase and tyrosine kinase etc.)⁷. We try to develop staurosporine analogs for new and novel type of anti-platelet aggregation agents. Especially, the obstacle of this development is its vasodilation activity which staurosporine naturally has. Therefore the high-specificity for platelet is demanded. We now describe the biological effect of conversion of γ -lactam into γ -lactone functionality of 1 and the key step of our synthesis of these compounds which involves a remarkably efficient reaction.

The analog (2), which was converted γ -lactam into γ -lactone of 1, was synthesized in good yield in four steps (Scheme 1). 1 has a methylamino functionality that may interfere with a series of reactions for 2. Therefore, its moiety was protected by 2,2,2-trichloroethoxycarbonyl (troc) group8. Treatment of troc-staurosporine (6) with DDQ/MeOH gave 7-methoxy product (7), which was hydrolyzed with HCl/H₂O-Dioxane to 7-hydroxy-troc-staurosporine (8). Treatment of 8 with NaBH₄ in Dioxane-aq.NaH₂BO₃ at 80°C afforded lactone (9), presumably via tautomerization from hydroxy- γ -lactam to amide-aldehyde, which was attacked at aldehyde carbon by nucleophile (H), then cyclization was occurred as shown in Equation 1. Then reduction with Zn/1N aq. NH₄OAc in THF provided deprotected γ -lactone (2).

Scheme 1: (a) Troc-Cl, Et₃N, CH₂Cl₂, 0°C~rt,92%; (b) DDQ, MeOH, CH₂Cl₂, rt, 96%; (c) 1N-HCl, H₂O/Dioxane, 60-65°C, 85%; (d) NaBH₄, Dioxane-1M aq. NaH₂BO₃, 80°C, 72%; (e) Zn, 1M-NH₄OAc(aq.), THF, rt, 72%.

Equation 1

Methylation of 2 with HCHO/NaBH₃CN gave 4'-N-Me-product (3). And reduction of 2 with LiAIH₄ in THF at room temperature gave diol (5) (Scheme 2).

Dihydroxy derivative (4) has been synthesized by the route shown in Scheme 3. Formylation of 9 with $Cl_2CHOCH_3/TiCl_4$ in dichloromethane⁹ gave the difformyl intermediate (1 1). Baeyer-Villiger oxidation of 1 1 with H_2O_2/H_2SO_4 in MeOH, followed by Zn-reduction afforded 4.

Scheme 2: (a) NaBH₃CN, 35% aq. HCHO, CH₃CN-DMF, rt, 79%; (b) LiAlH₄, THF, rt, 80%.

Scheme 3: (a) Cl₂CHOMe, TiCl₄, CH₂Cl₂, -20°C~rt, 71%; (b) H₂O₂, H₂SO₄, MeOH, rt, 32%; (c) Zn, 1M aq. NH₄OAc, THF, reflux,72%.

The analogs prepared using above methodology showed potent inhibition activity against platelet aggregation. Aggregation of platelet was measured using a NBS hematracer VI (Nikoo Bioscience Co. Ltd., Japan). Washed platelet was prepared from guinea pig blood by gel filtration method 10 . An aliquot of washed platelet $(6x10^8 \text{ cells/ml})$ was incubated with various concentration of inhibitors for 3 minutes and stimulated with $^{1}\mu\text{M}$ U46619 for 10 minutes.

Table. Inhibitory effect arterial contraction, platelet aggregation and PKC activity

Compound No	IC50 values (μM)		
	Arterial contraction	Platelet aggregation	PKC activity
1	0.025	7.5	0.023
2	>30	6.8	5.8
3	>10	5.4	56
4	3.0	0.4	0.22
5	30	6.8	15.8

The IC₅₀ values against platelet aggregation was $6.8\mu M$ for 2, which was much the same as compared with that of staurosporine (7.5 μM). On the other hand, the IC₅₀¹¹ values against smooth muscle contraction was > $30\mu M$ for 2, which was much higher than that of staurosporine (0.025 μM). Thus, the γ -lactam portion of 1 could be converted into γ -lactone such as 2 without loss of antiplatelet aggregation activity. Furthermore, 2 did not show inhibition activity against smooth muscle contraction. These results suggested that conversion of γ -lactam of staurosporine (1) into γ -lactone plays an important role to show platelet-selectivity. 4'-N-Methyl derivative (3) and diol (5) showed the same antiplatelet activity (IC₅₀: 5.4 μM and 6.8 μM , respectively) as compared with 2. Both modification of sugar moiety and ring-opening of γ -lactone may not hve much effective for anti-platelet activity. Compound (4), which was substituted with hydroxy groups on aromatic portion of 2, was improved for antiplatelet aggregation activity about by 10 times (IC₅₀: 0.4 μM) than 2. Substitution on aromatic portion may be important for augmentation of antiplatelet aggregation activity. The IC₅₀¹¹ for PKC,

which contains various types of isozymes, were 23nM for 1 and 220nM for 4. Accordingly, PKC may not be important for platelet aggregation with respect to these compounds. Kinases other than PKC, probably, reflect on platelet aggregation.

References and Notes:

- 1. Ōmura, S.; Iwai, Y.; Hirano, A.; Nakagawa, A.; Awaya, J.; Tsuchiya, H.; Takahashi, Y.; Masuma, R. J. Antibiotics 1977, 30, 275.
- 2. Tamaoki, T.; Nomoto, H.; Takahashi, I.; Kato, Y.; Morimoto, M.; Tomita, F. *Biochem. Biophys. Res. Commun.* **1986**, *135*, 397.
- 3. Omura, S.; Iwai, Y. Hakko to Kougyo 1979, 37, 223.
- 4. Sasaki, Y.; Seto, M.; Komatsu, K.; Omura, S. Eur. J. Pharmacol. 1991, 202, 367.
- 5. Okazaki, T.; Kato, Y.; Mochizuki, T.; Tashima, M.; Sawada, H.; Uchino, H. *Exp. Hematol.* **1988**, *16*, 42.
- 6. Matsumoto, H.; Sasaki, Y. Biochem. Biophys. Res. Commun. 1989, 158, 105.
- 7. Ōmura, S.; Sasaki, Y.; Iwai, Y.; Takeshima, H. J. Antibiotics 1995, 48, 535.
- 8. Windholz, T. B.; Johnston, D. B. R. Tetrahedron Lett. 1967, 27, 2555.
- 9. Magnus, P. D.; Sear, N. L. J. Org. Chem. 1973, 38, 4243.
- 10. Tangen, O.; Berman, H. J.; Marfey, P. Tromb. Diath. Haemorrth. 1971, 25, 269.
- 11. Asano, M.; Matsunaga, K.; Miura, M.; Ito, K. M.; Seto, M.; Sakurada, K.; Nagumo, H.; Sasaki, S.; Ito. K. Eur. J. Pharmacol. 1995, 294, 693.

(Received in Japan 8 May 1996; accepted 10 July 1996)